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SYNTHESIS AND APPLICATION OF N(6)-PHENOXYCARBONYL-DEOXY-ADENOSINE DERIVATIVES IN OLIGONUCLEOTIDE PROBES CHEMISTRY

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<u>Abstract:</u> N(6)-phenoxycarbonyl-deoxyadenosine derivatives are offered as new reactive intermediates for preparation of linker-containing oligonucleotide probes and modified LCAA-CPG supports.

During last years much effort has been undertaken in construction of oligonucleotide probes containing nucleoside units bearing reactive linkers, mainly of alkylamine type /1-3/. Here we would like to communicate on the synthesis and application of N(6)-phenoxycarbonyl-deoxyadenosine derivatives in oligonucleotide probes chemistry.

Deoxyadenosine was 5'-O-dimethoxytritylated (70% yield) with use of 6-nitroquinoline as a base /4/. Under such conditions no tritylation of exo-amine function was observed. Subsequent 3'-O-silylation with tBDMSi-chloride (dioxane-imidazole, 4 days,room temp. 95% yield) and treatment with crystalline phenoxycarbonyltetrazole /5/ led quantitatively to the 3',5'-di-O-protected-N(6)-phenoxycarbonyldeoxyadenosine $\underline{1}$ /6/ (85%) as a highly reactive intermediate /7,8/. Reaction of the latter with 1,6-di-aminohexane gave (80%) a derivative bearing alkylamine-chain linked to nebularin-6-yl residue via, ammonia-resistant, N^1 , N^2 -disubstituted urea system /9/. Hexylamine protection with trifluoroacetic anhydride (70%), removal of 3'-O-TBDMSi group with $\text{Et}_3 \, \text{NH}^+ \, \text{F}^-$ (94%) and subsequent treatment with bis-(N,N-diiso-propylamino)(2-cyanoethoxy)phosphine gave (as lyophilisate, 95% yield) requested phosphoramidite $\underline{2}$ /10/.

DMTr0
$$\frac{1}{1}$$
 DMTr0 $\frac{1}{1}$ $\frac{1}{1}$ CNCH2CH20 $\frac{1}{1}$ $\frac{$

Phosphoramidite 2 was applied for automated DNA synthesis to obtain precursor-oligonucleotides bearing free aminohexyl linker capable to react with reagents introducing reporter probes /3/. Below are examples of oligeoxyonucleotide probes obtained and HPLC-purified:

- Fluorescein-linker-N(CAC CAC CAC CAC CAC); DNA-fingerprinting probe

- Biotin-linker-N(GTA AAA CGA CGG CCA GT) ; M13-sequencing primer

- thiol-linker-N(CCT AGT GGA GGA AAG A) ; 16s rRNA probe

N(6)-phenoxycarbonyldeoxyadenosine derivatives e.g. $\underline{1}$ react readily with LCAA-CPG (loading 32 μ mole/g) to obtain anchored (ammonia-stable) CPG-linked oligonucleotides. When appropriate linker is to be derived from threonine, formation of detachable (ammonia labile) CPG-linked oligonucleotide is expected as in the case of hypermodified ureidonucle-side t⁶ A /11/. Potential application of the latter approach to the solid support-aided synthesis of RNA-branch is investigated.

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- 6. 1 was isolated by flash column chromatography on silica gel using methanol gradient in chloroform or used in situ after careful hydrolysis of an excess of phenoxycarbonyltetrazole as described /7/; characterized via its 3',5'-di-O-acetyl derivative, compare /5/, H NMR (90 MHz, CDCl3); δ(ppm) 9.31(1H,s,N6-H,rapid exchange in D20),8.90(1H,s,H-8),8.30(1H,s,H-2),7.50(5H,m,phenyl),6.40(t,1,H-1').
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- 9. reaction of 3'-0-acetyl analogue with conc.ammonia $(50^{\circ}\text{C},48\text{hrs})$ led only to removal of 3'-0-protection, showing stability of N1,N2-disubstituted urea system, easily detectable due to characteristic λ max at 300 nm in a basic solution (pH 12.5).
- 2 H NMR (90 MHz, CDC13); δ(ppm) 10.30(1H,m,N-H,exchange in D20), 9,40(1H,s,N6-H,rapid exchange in D20), 8.67(1H,s,H-8), 8.20 (1H,s,H-2), 6.37(1H,t,H-1'), 3.75(6H,s,Tr-OMe), 1.44-1.36(12H,br-m hexyl), 0.88(15,br-s,tBDMSi); P31 NMR (32.6 MHz, ext.85% H3PO4) δ(ppm) 148.43.
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